UDC 544.723

doi: 10.15407/hftp15.03.429

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CAFFEIC ACID COMPLEXES OVER ALUMINA SURFACE: FTIR, TPD MS AND DFT CALCULATIONS

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Caffeic acid (CA) is a phenolic, natural, biologically active compound with pronounced antioxidant and antimicrobial properties. It has a great potential for use in medicine and cosmetology and can also be used to obtain a number of other useful chemicals. Therefore, research aimed at improving the technologies for removing caffeic acid from plant raw materials and its processing is relevant. In our work, we have studied the complexes of caffeic acid with nanosized Al_2O_3 , which is usually used in various plant biomass conversion technologies, using FT-IR spectroscopy, temperature-programmed desorption mass spectrometry (TPD MS), and DFT calculations. The FT-IR spectra analysis shows that CA can interact with aluminum oxide via both the carboxyl and phenolic groups. Based on the measured differences between carbonyl symmetric and asymmetric vibrational peaks (Δv), it was found that carboxylate complexes of CA on the Al_2O_3 surface can have bidentate and monodentate structures. The mass spectrometric data analysis made it possible to identify compounds of 4-vinyl catechol, pyrocatechol, and phenol, which are decomposition products of the formed carboxylate and phenolic complexes. It was found that on the surface of the studied CA/Al₂O₃ samples, CA chelate complexes, formed with the participation of both OH groups of the aromatic ring, predominate.

Keywords: biomass, carboxylate complexes, chelate complexes, 4-vinyl catechol, pyrocatechol, phenol, pyrolysis, biomass conversion technologies

INTRODUCTION

Caffeic acid is a C6-C3 phenolic natural biologically active compound, which has pronounced antioxidant, antimicrobial and other biological activities [1–5], and has great potential for use in medicine [6, 7]. It is often found in plants that are used in food, and is mainly found in them as ester or bound to biopolymers [8]. CA, both free and esterified, is usually the most abundant phenolic acid and accounts for 75 to 100 % of the total hydroxycinnamic acid content in most fruits [9]. It is mainly involved in the synthesis of lignin, as well as the regulation of cell expansion, turgor pressure, phototropism, water flow and growth [10]. At the same time, a significant part of the biomass, which is the waste of the agroindustrial complex, can be used to obtain caffeic acid for use in medicine, cosmetology and other areas. In particular, the source of CA can be pulp of apples [12–15], grapes [15–19], tomatoes etc. [20, 21], which are by-products of juice

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Aluminum oxide is one of the oxides commonly used as a catalyst and carrier, including in various plant biomass conversion technologies [38-41]. Therefore, the study of caffeic acid complexes, which they form when interacting with aluminum oxide, is significant for the development and optimization of CA-obtaining technologies, as well as technologies for processing plant raw materials into various chemicals that involve the use of aluminum oxide. In particular, data on the type of complexes formed and their relative amount on the oxide surface combined with data on their stability, as well as an understanding of the mechanisms of the biomass components transformations under the influence of various physical factors (irradiation, heating, solvents, and others) [11, 19, 20, 25, 23, 29, 32, 33], will allow determining the possibilities of using Al₂O₃ in those or other technologies.

our work is devoted Therefore. to determining the complexes formed by caffeic acid with nanosized aluminum oxide using FT-IR spectroscopy, temperature-programmed desorption mass spectrometry, and DFT calculations.

EXPERIMENTAL

The research used caffeic acid (≥ 98 %, Sigma-Aldrich) and alumina (99.5 %. $S_{Ar} = 89 \text{ m}^2/\text{g}$, Evonic), which had previously been calcined at 500 °C for two hours to remove any organic based impurities.

 CA/Al_2O_3 samples (0.3 and 0.6 mmol/g) were prepared by impregnating 100 mg of aluminum oxide with 2 ml of a solution of caffeic acid in ethanol (96%). The resulting suspensions were mixed and dried in the open air at room temperature.

IR-spectral studies were performed using a Fourier transform Thermo Nicolet NEXUS device in the diffuse reflectance mode. For research, samples of CA/Al₂O₃ and pure Al₂O₃ were mixed with KBr in a ratio of 1:10, and pure CA - in a ratio of 1:100. All samples were thoroughly ground for the same amount of time. The spectra were recorded with a resolution of 4 cm^{-1} . The scanning speed was 0.5 cm/s, and the number of scans was 50.

Examination of the samples using the method of temperature-programmed desorption mass spectrometry (TPD MS) was performed on a monopole mass spectrometer MH-7304A (Sumy, Ukraine) with electron ionization, converted for conducting thermodesorption measurements [42]. A sample weighing 10-20 mg was placed in a quartz-molybdenum ampoule and pumped at room temperature to a pressure of ~ $5 \cdot 10^{-5}$ Pa. The sample was heated at the rate of 0.17 °C/s from room temperature to 750 °C. Volatile decomposition products entered the ionization chamber of the mass spectrometer, where they were ionized and fragmented under the influence of electrons. The intensities of the ion currents of the products of desorption and thermolysis in the mass analyzer were recorded by a secondary electron multiplier BEU-6. The range of investigated masses was m/z 1–220. Registration and analysis of mass spectra were carried out by an automated computer system.

Density functional (DFT) calculations were performed using the wB97XD functional with Grimme's D2 dispersion corrections [43] and the 6-311++G(d,p) basis set. Gaussian 09 software version B01 [44] was used. For the studied molecular systems, full geometry optimization was performed. The total charge of the system was zero. The results of the calculations were visualized using the molecular graphics program MaSK v. 1.3.0 [45]. As the simplest model of aluminum oxide, a cluster of composition $Al_2O_6H_6$ [46] was used.

RESULTS AND DISCUSSION

FT-IR spectroscopic study. To find out which functional groups of CA are involved in the interaction with surface of aluminum oxide, a spectroscopic study of the CA/Al₂O₃ sample (0.3 mmol/g) was carried out by the FT-IR method (Fig. 1).

For the CA/Al₂O₃ sample, the absence of an absorption maximum at 1645 cm⁻¹, which corresponds to valence vibrations of the carbonyl group (v(C=O)) [47], was found. This indicates the formation of carboxylate complexes. The new broad band at 1630 cm⁻¹, which was masked by intense absorption of v(C=O) in the CA spectrum, probably refers to v(C=C)vibrations. A shoulder appears around 1670 cm⁻¹ for CA/Al₂O₃, which may correspond to v(C=O). Whereas the absorption detected in the spectra of the studied samples in the region of 1394 cm⁻¹ probably corresponds to v(CO) vibrations. It has been demonstrated that the magnitude of " Δv " the separation between the asymmetric and symmetric carboxylate stretches ($\Delta v = v_{as}(COO^{-})$ or, in case of monodentate $v_s(COO^-))$ coordination, between the C=O and C-O stretches ($\Delta v = v(C=O) - v(CO)$) can be used to determine the type of the carboxylate (and nondissociated acid) binding [48, 49].

The absorptions at 1670 cm⁻¹ ν (C=O) and 1394 cm⁻¹ v(\overline{CO}) most likely correspond to monodentate complexes, since the value of Δv is 276 cm⁻¹. The assignment of absorption bands to mono- and bidentate complexes was previously made on the example of surface complexes of valeric acid on the surface of metal oxide catalysts (SiO₂, y-Al₂O₃, CeO₂/SiO₂, Al₂O₃/SiO₂ and TiO₂/SiO₂) based on the values of Δv [28]. The results of quantum chemical calculations and IR spectroscopy [50] supported such an interpretation. According to [50], the absorption at ~ 1680 cm^{-1} detected for valeric acid corresponds to monodentate carboxylate complexes. In addition, the CA/Al₂O₃ spectra contain absorptions in the region of 1400 and 1568 cm^{-1} , which apparently correspond to



Fig. 1. FT-IR spectra in the regions $1100-1800 \text{ cm}^{-1}$ and $2000-3800 \text{ cm}^{-1}$: (a) pure Al₂O₃, (b) CA/Al₂O₃, (c) pure CA

Some changes were also detected in the region of valence vibrations of OH groups (Fig. 1 b). In particular, the maxima at 3226 and 3440 cm^{-1} are not observed. There is only a broad band with a maximum of 3421 cm^{-1} . These bands belong to the valence vibrations of phenolic hydroxyls [47, 48]. The first maximum may correspond to phenolic groups forming intermolecular hydrogen bonds in CA associates. The maximum at 3440 cm^{-1} is in the region where intermolecular hydrogen bonds in dimers of CA and intramolecular hydrogen bonds of CA are manifested [49]. Although the absorption of hydrogen-bonded OH groups of water, which may be present in the studied samples, may also appear here. The absence of a band at 3226 cm⁻¹ indicates the destruction of intermolecular hydrogen bonds between phenolic groups.

In addition, the part of the spectrum between $1000-1500 \text{ cm}^{-1}$ changes significantly for the CA/Al₂O₃ sample. Absorptions of COH-groups for phenolic and carboxyl groups appear here, so identifying bands in this part of the spectrum is challenging. The analysis of this area was performed based on a number of our own FT-IR spectroscopic studies of phenolic compounds [53, 54] and literature data [47, 55–57]. According to [47, 55], a broad intense band located around 1300 cm⁻¹, which often has several maxima, is due to the superimposition of COH vibrations of phenolic and carboxylic OH groups. For the CA/Al₂O₃ sample (0.3 mmol/g),

this band undergoes a transformation, in particular, its maximum shifts from 1282 to 1269 cm⁻¹. The peak at 1298 cm⁻¹, which may correspond to plane deformation vibrations of phenolic OH groups [47], practically disappears. In addition, the 1218 cm^{-1} band, which is also attributed to COH vibrations [47], in the CA/Al_2O_3 spectrum (0.3 mmol/g) has a much lower relative intensity than for CA. The bands at 1327 and 1352 cm⁻¹ disappear, which can be manifested as plane deformation vibrations of OH groups [52]. Instead, absorption appears at 1338 cm⁻¹. Such changes may mean that not only the carboxyl OH group, but also phenolic groups participate in the interaction with the oxide. According to a number of literature data [58-60], the formation of CA complexes with the participation of phenolic groups can also be evidenced by the position of the two most intense bands in the CA/Al_2O_3 (0.3 mmol/g) spectrum - near 1269 and 1497 cm⁻¹. At the same time, the first of them is attributed to the C-OH valence vibrations of the aromatic nucleus, and the second to the v(C=C) vibrations of the benzene ring. This spectrum characteristic is a sign of chelate complexes forming phenolic groups of caffeic acid with various oxides [58-60].

DFT calculations. We calculated the possible structures of carboxylate complexes of caffeic acid on the aluminum oxide surface. Table 1 shows the value of the Gibbs free energy

symmetric ($v_s(COO^-)$) and asymmetric ($v_{as}(COO^-)$) valence vibrations of carboxylate complexes. The structure of these complexes is bidentate, since Δv is 168 cm⁻¹.

at the temperature of 421 K (G421) for the calculated structures. The structures of CA complexes with a cluster of composition $Al_2O_6H_6$ are presented in Fig. 2.

According to the performed calculations, caffeic acid can form phenolate complexes (F1 and F2) and both monodentate carboxylate (C1-C3) and bidentate bridge (C4) complexes over alumina, Fig. 2.



Fig. 2. Possible structures of caffeic acid complexes with alumina (carboxylate complexes: C1, C2, C3, C4 and phenolate complexes – F1, F2)

 Table 1. Gibbs free energy values for the temperature 421K (G421) for the calculated structures of caffeic acid complexes over alumina

	G ₄₂₁ , a.u.				
C1	C2	C3	C4	F 1	F2
-1588.771629	-1588.778037	-1588.773954	-1588.774137	-1588.787403	-1588.786720

Study of the CA/Al_2O_3 sample by the TPD MS. The pyrolysis of CA occurred in the temperature range from 100 to 600 °C and was

accompanied by the formation of a large number of products with m/z from 3 to 178, which can be seen in the mass spectra obtained during the TPDMS study of the sample of CA/Al₂O₃ (0.6 mmol/g) (Fig. 3). The main products of CA decomposition on the Al₂O₃ surface included: 4-vinyl catechol (*m/z* 136, 110–400 °C), catechol (m/z 110, $T_{\text{max}} = 123 \text{ °C}$), phenol (m/z, 94, $T_{\rm max} = 409 \,^{\circ}{\rm C}$ and others. In the process of pyrolysis, H₂O (m/z 18), CO₂ (m/z 44), and CO $(m/z \ 28)$ were released intensively. Based on the analysis of the obtained TPD MS data (Fig. 3) and the results of studies of the thermal decomposition of cinnamic acids on the surface of ceria [53], certain conclusions can be drawn about CA complexes that form on the aluminum oxide surface.

From the results [53, 54, 61], it is clear that the destruction of carboxylate complexes of cinnamic acids on the CeO₂ occurs mainly by decarboxylation and the formation of the corresponding vinyl phenol (styrene). At the same time, it is shown that decarboxylation of various types of carboxylate complexes occurs at different temperatures. It is known that free phenolic acids, particularly caffeic, undergo decarboxylation both under the influence of temperatures [62] and in the metabolism processes of various microorganisms [63].

As shown in Figs. 3 and 4, 4-vinyl catechol (M.r. = 136 Da, m/z 136) belongs to the main decomposition product of CA pyrolysis over alumina. Its formation was registered in a wide range of temperatures (110-400 °C). Thus, its release is probably related to the destruction of various types of carboxylate complexes and associates of CA, which are formed when the CA amount increases on the surface of the oxide, whereas catechol (m/z 110) and phenol (m/z 94) (Figs. 3, 4) can be products of the destruction of complexes formed with the participation of phenolic OH groups [53]. The first compound may be a product of the decomposition of CA molecules bound to the Al₂O₃ surface through one aromatic hydroxyl. Phenol is probably released during the destruction of chelate complexes formed due to the interaction of both phenolic groups of CA with the oxide surface. Thus, the registration of these compounds in the mass spectra of CA/Al₂O₃ during heating can be a sign of the presence of a number of carboxylate and phenolic complexes on the Al₂O₃ surface, which was also revealed by FT-IR spectroscopic data.



Fig. 3. Mass spectra of pyrolysis products of the CA/Al₂O₃ sample, obtained at temperatures of 194 °C (*a*) and 400 °C (*b*)



Fig. 4. TPD curves of molecular ions of (*a*) vinyl catechol (m/z 136, curve 1), catechol (m/z 110, curve 2); and (*b*) phenol (m/z 94), obtained during pyrolysis of the CA/Al₂O₃ sample

At the same time, the high relative release intensity of phenol (m/z 94) compared to catechol (m/z 110) and vinyl catechol (m/z 136)may indicate a significant content of complexes that are formed with the participation of both phenolic groups. This is consistent with the results of the FT-IR spectroscopic study. As shown in Fig. 1, the relative intensity of the carboxylate bands is relatively low. In addition, it is known that caffeic acid, which has osubstituted hydroxyl groups in its structure, can form chelate phenolic complexes with metals [60, 64, 65].

CONCLUSIONS

Samples of CA over the alumina surface were investigated using FT-IR spectroscopy, TPD MS, and quantum chemical methods. From the obtained FT-IR spectroscopic data, it was found that the interaction of CA with aluminum oxide takes place with the participation of both carboxyl and phenolic groups. The formation of such products as 4-vinyl catechol (M.r. = 136 Da,

m/z 136), catechol (M.r. = 110 Da, m/z 110) and phenol (M.r. = 94 Da, m/z 94), which were registered by TPD MS in the study of CA/Al₂O₃ samples, was associated with the destruction of a number of carboxylate and phenol complexes of CA. This can serve as confirmation of the presence of these complexes on the Al₂O₃ surface. The obtained data indicate that phenolic complexes predominate for the studied samples.

ACKNOWLEDGEMENTS

This research has received funding through the EURIZON project, which is funded by the Union European under grant agreement No. 871072. This work was partially supported by the Cardiff HEFCW ODA grant, the grant FSA3-20-66700 from the U.S. Civilian Research & Development Foundation (CRDF Global) with funding from the United States Department of and the Simons Support State, Grant No. 1290589. We would also like to thank the Armed Forces of Ukraine for providing security to perform this work.

Комплекси кавової кислоти на поверхні оксиду алюмінію: ІЧ спектроскопія, ТПД МС та DFT розрахунки

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Кавова кислота є фенольною природною біологічно активною сполукою з вираженими антиоксидантними та антимікробними властивостями, яка має великий потенціал застосування в медицині і косметології, а також може використовуватися для отримання ряду інших корисних хімікатів. Тому дослідження, спрямовані на вдосконалення технологій вилучення кавової кислоти з рослинної сировини, а також її переробки, є актуальними. В нашій роботі вивчалися комплекси кавової кислоти з нанорозмірним Al₂O₃, який зазвичай застосовують в різних технологіях конверсії рослинної біомаси. Структуру та тип утворених комплексів СА досліджено за допомогою ІЧ-спектроскопії, температурно-програмованої десорбційної мас-спектрометрії (ТПД МС) та квантовохімічних методів. Аналіз отриманих ІЧ-спектрів свідчить, що СА може взаємодіяти з нанорозмірним оксидом алюмінію як карбоксильною так і фенольними групами. На основі розрахованих значень Δv встановлено, що карбоксилатні комплекси СА на поверхні Al₂O₃ можуть мати бідентатну та монодентатну структури. Аналіз мас-спектрометричних даних дозволив ідентифікувати сполуки 4-вінілкатехолу, пірокатехолу та фенолу, які є продуктами розкладу утворених карбоксилатних та фенольних комплексів. Виявлено, що на поверхні досліджених зразків CA/Al₂O₃ переважають хелатні комплекси СА, які утворюються за участі обох OH-груп ароматичного кільця.

Ключові слова: біомаса, карбоксилатні комплекси, хелатні комплекси, 4-вінілкатехол, пірокатехол, фенол, піроліз, технології конверсії біомаси

REFERENCES

- 1. Stojković D., Petrović J., Soković M., Glamočlija J., Kukić-Marković J., Petrović S. *In situ* antioxidant and antimicrobial activities of naturally occurring caffeic acid, p-coumaric acid and rutin, using food systems. *J. Sci. Food Agric*. 2013. **93**(13): 3205.
- Zhaveh S., Mohsenifar A., Beiki M., Khalili St., Abdollahi A., Rahmani-Cherati T., Tabatabaei M. Encapsulation of Cuminum cyminum essential oils in chitosan-caffeic acid nanogel with enhanced antimicrobial activity against Aspergillus flavus. *Ind. Crops Prod.* 2015. 69: 251.
- Anwar J., Spanevello R.M., Thomé G., Stefanello N., Schmatz R., Gutierres, J., Vieira J., Jucimara Baldissarelli, Carvalho F.B., da Rosa M.M., Rubin M.A., Fiorenza A., Morsch V.M., Schetinger M.R.C. Effects of caffeic acid on behavioral parameters and on the activity of acetylcholinesterase in different tissues from adult rats. *Pharmacol. Biochem. Behav.* 2012. **103**(2): 386.
- 4. Khan F., Bamunuarachchi N.I., Tabassum, N., Kim Y.M. Caffeic acid and its derivatives: antimicrobial drugs toward microbial pathogens. *J. Agric. Food Chem.* 2021. **69**(10): 2979.
- Lima V.N., Oliveira-Tintino C.D., Santos E.S., Morais L.P., Tintino S.R., Freitas T.S., Geraldo Y.S., Pereira R.L.S., Cruz R.P., Menezes I.R.A., Coutinho H.D. Antimicrobial and enhancement of the antibiotic activity by phenolic compounds: Gallic acid, caffeic acid and pyrogallol. *Microb. Pathogen.* 2016. 99: 56.
- 6. Russo G.I., Campisi D., Di Mauro M., Regis F., Reale G., Marranzano M., Morgia G. Dietary consumption of phenolic acids and prostate cancer: A case-control study in sicily, Southern Italy. *Molecules*. 2017. **22**(12): 2159.
- Muhammad Abdul Kadar N.N., Ahmad F., Teoh S.L., Yahaya M.F. Caffeic acid on metabolic syndrome: a review. *Molecules*. 2021. 26(18): 5490.
- 8. Clifford M.N. Chlorogenic acids and other cinnamates-nature, occurrence and dietary burden. J. Sci. Food Agric. 1999. **79**(3): 362.

- 9. Manach C., Scalbert A., Morand C., Rémésy C., Jiménez L. Polyphenols: food sources and bioavailability. *Am. J. Clin. Nutr.* 2004. **79**(5): 727.
- 10. Lattanzio V., Lattanzio V.M., Cardinali A. Role of phenolics in the resistance mechanisms of plants against fungal pathogens and insects. *Phytochemistry: Advances in Research*. 2006. **661**(2): 23.
- 11. Qian S., Lu M., Zhou X., Sun S., Han Z., Song, H. Improvement in caffeic acid and ferulic acid extraction by oscillation-assisted mild hydrothermal pretreatment from sorghum straws. *Bioresour. Technol.* 2024. **396**: 130442.
- 12. Chandrasekar V., Martín-González M.S., Hirst P., Ballard T.S. Optimizing Microwave-Assisted Extraction of Phenolic Antioxidants from Red Delicious and Jonathan Apple Pomace. J. Food Process Eng. 2015. **38**(6): 571.
- 13. Bai X.L., Yue T.L., Yuan Y.H., Zhang H.W. Optimization of microwave-assisted extraction of polyphenols from apple pomace using response surface methodology and HPLC analysis. *J. Sep. Sci.* 2010. **33**(23–24): 3751.
- 14. Zhang M., Wang D., Gao X., Yue Z., Zhou H. Exogenous caffeic acid and epicatechin enhance resistance against Botrytis cinerea through activation of the phenylpropanoid pathway in apples. *Sci. Hortic.* 2020. **268**: 109348.
- 15. Lee J., Chan B.L.S., Mitchell A.E. Identification/quantification of free and bound phenolic acids in peel and pulp of apples (Malus domestica) using high resolution mass spectrometry (HRMS). *Food Chem.* 2017. **215**: 301.
- Casagrande M., Zanela J., Pereira D., de Lima V.A., Oldoni T.L.C., Carpes S.T. Optimization of the extraction of antioxidant phenolic compounds from grape pomace using response surface methodology. *J. Food Meas. Charact.* 2019. 13: 1120.
- Tournour H.H., Segundo M.A., Magalhaes L.M., Barreiros L., Queiroz J., Cunha L.M. Valorization of grape pomace: Extraction of bioactive phenolics with antioxidant properties. *Ind. Crops Prod.* 2015. 74: 397.
- Otero-Pareja M.J., Casas L., Fernández-Ponce M.T., Mantell C., Martinez de la Ossa E.J. Green extraction of antioxidants from different varieties of red grape pomace. *Molecules*. 2015. 20(6): 9686.
- 19. Fracassetti D., Lawrence N., Tredoux A.G.J., Tirelli A., Nieuwoudt H.H., Du Toit W.J. Quantification of glutathione, catechin and caffeic acid in grape juice and wine by a novel ultra-performance liquid chromatography method. *Food Chem.* 2011. **128**(4): 1136.
- 20. Vorobyova V., Skiba M., Vasyliev G. Extraction of phenolic compounds from tomato pomace using choline chloride–based deep eutectic solvents. *J. Food Meas. Charact.* 2022. **16**(2): 1087.
- 21. Farinon B., Felli M., Sulli M., Diretto G., Savatin D.V., Mazzucat A., Costantini L. Tomato pomace food waste from different variants as a high antioxidant potential resource. *Food Chem.* 2024. **452**: 139509.
- 22. Hu M., Zhu G., Chen Y., Xie G., Zhu M., Lv T., Xu L. Enhanced co-pyrolysis of corn stalk and bio-tar into phenolicrich biooil: Kinetic analysis and product distributions. *J. Anal. Appl. Pyrolysis.* 2024. **177**: 106358.
- 23. Pereira P.H., Maya D.M., Oliveira D.C., Ferreira A.F. From waste to resource: maximizing olive pomace valorization through advanced thermal treatment. *Biomass Convers. Biorefin.* 2024. 1-17.
- 24. Velvizhi G., Jacqueline P.J., Shetti N.P., Latha K., Mohanakrishna G., Aminabhavi T.M. Emerging trends and advances in valorization of lignocellulosic biomass to biofuels. *J. Environ. Manag.* 2023. **345**: 118527.
- Nastasiienko N., Kulik T., Palianytsia B., Larsson M., Kartel M. Microwave-assisted catalytic pyrolysis of ferulic acid, as a lignin model compound. *J. Therm. Anal. Calorim.* 2023. 148(12): 5485.
- Sankaranarayanan S., Won W. Catalytic pyrolysis of biomass to produce bio-oil using layered double hydroxides (LDH)-derived materials. GCB Bioenergy. 2024. 16(3): e13124.
- Sarkar R., Laskar N., Saha A., Basak B.B. Green biorefinery for residual biomass from agriculture. In: *Plant Biomass Applications*. (Academic Press, 2024).
- Fu W., Bai X., Tursun Y., Liu Q., Li B., Dai Z., Zhao Y., Li X., Guo L., Li J. Oxidative pyrolysis of plywood waste: Effect of oxygen concentration and other parameters on product yield and composition. *J. Anal. Appl. Pyrolysis.* 2023. **173**: 106068.
- 29. Chaabane A., Abderafi S., Abbassi M.A. Valorizing argan residues into biofuels and chemicals through slow pyrolysis. *Results Eng.* 2024. **21**: 101659.
- Du J., Shen T., Hu J., Zhang F., Yang S., Liu H., Wang H. Study on thermochemical conversion of triglyceride biomass catalyzed by biochar catalyst. *Energy*. 2023. 277: 127733.
- Tran M.H., Paramasivam P., Le H.C., Nguyen D.T. Biomass: A Versatile Resource for Biofuel, Industrial, and Environmental Solution. *International Journal on Advanced Science, Engineering and Information Technology*. 2024. 14(1): 17489.
- 32. Quintero-Naucil M., Salcedo-Mendoza J., Solarte-Toro J.C., Aristizábal-Marulanda V. Assessment and comparison of thermochemical pathways for the rice residues valorization: pyrolysis and gasification. *Environ. Sci. Pollut. Res.* 2024. 1.
- Wang B., Chen Y., Chen W., Hu J., Chang C., Pang S., Li P. Enhancement of aromatics and syngas production by co-pyrolysis of biomass and plastic waste using biochar-based catalysts in microwave field. *Energy*. 2024. 293: 130711.
- Len T., Bressi V., Balu A.M., Kulik T., Korchuganova O., Palianytsia B., Esprob C., Luque R. Thermokinetics of production of biochar from crop residues: an overview. *Green Chem.* 2022. 24(20): 7801.

- Yefremova S., Zharmenov A., Sukharnikov Y., Bunchuk L., Kablanbekov A., Anarbekov K., Kulik T., Nikolaichuk A., Palianytsia B. Rice husk hydrolytic lignin transformation in carbonization process. *Molecules*. 2019. 24(17): 3075.
- 36. Shafizadeh A., Rastegari H., Shahbeik H., Mobli H., Pan J., Peng W., Li G., Tabatabaei M., Aghbashlo M. A critical review of the use of nanomaterials in the biomass pyrolysis process. *J. Cleaner Prod.* 2023. **400**: 136705.
- Hu M., Zhu G., Chen Y., Xie G., Zhu M., Lv, T., Xu L. Enhanced co-pyrolysis of corn stalk and bio-tar into phenolicrich biooil: Kinetic analysis and product distributions. J. Anal. Appl. Pyrolysis. 2024. 177: 106358.
- Liu W., Liu B., Zhang Y., Yi B., Hu H., Fan Q., Liu H. Evolution of Pyrolysis Characteristics and Gas Components of Biochar Prepared by Either Mixing or Layering Rice Husk with Inert Aluminum Oxide. *BioResources*. 2023. 18(1). 10.
- Pryhunova O., Dyachenko A., Ischenko O., Diuyk V., Goncharuk O., Oranska O., Bonarowska M. NiFe (CoFe)/silica and NiFe (CoFe)/alumina nanocomposites for the catalytic hydrogenation of CO₂. *Appl. Nanosci.* 2023. 13(10): 6829.
- 40. Qasemi Z., Jafari D., Jafari K., Esmaeili H. Heterogeneous aluminum oxide/calcium oxide catalyzed transesterification of Mespilus germanica triglyceride for biodiesel production. *Environ. Prog. Sustainable Energy*. 2022. **41**(2): e13738.
- 41. Sharanda L.F., Shimansky A.P., Kulik T.V., Chuiko A.A. Study of acid-base surface properties of pyrogenic y-aluminium oxide. *Colloids Surfaces A*. 1995. **105**(2–3): 167.
- 42. Kulik T., Palianytsia B., Larsson M. Catalytic pyrolysis of aliphatic carboxylic acids into symmetric ketones over ceria-based catalysts: kinetics, isotope effect and mechanism. *Catalysts*. 2020. **10**(2): 179.
- 43. Chai J.D., Head-Gordon M. Long-range corrected hybrid density functionals with damped atom-atom dispersion corrections. *Phys. Chem. Chem. Phys.* 2008. **10**: 6615.
- 44. Frisch M.J., Trucks G.W., Schlegel H.B., Scuseria G.E., Robb M.A., Cheeseman J.R., Scalmani G., Barone V., Petersson G.A., Nakatsuji H., Li X., Caricato M., Marenich A., Bloino J., Janesko B.G., Gomperts R., Mennucci B., Hratchian H.P., Ortiz J.V., Izmaylov A.F., Sonnenberg J.L., Williams-Young D., Ding F., Lipparini F., Egidi F., Goings J., Peng B., Petrone A., Henderson T., Ranasinghe D., Zakrzewski V.G., Gao J., Rega N., Zheng G., Liang W., Hada M., Ehara M., Toyota K., Fukuda R., Hasegawa J., Ishida M., Nakajima T., Honda Y., Kitao O., Nakai H., Vreven T., Throssell K., Montgomery J.A., Jr., Peralta J.E., Ogliaro F., Bearpark M., Heyd J.J., Brothers E., Kudin K.N., Staroverov V.N., Keith T., Kobayashi R., Normand J., Raghavachari K., Rendell A., Burant J.C., Iyengar S.S., Tomasi J., Cossi M., Millam J.M., Klene M., Adamo C., Cammi R., Ochterski J.W., Martin R.L., Morokuma K., Farkas O., Foresman J.B., Fox D.J.Gaussian 09, Revision B.01; Gaussian, Inc.: Wallingford, CT, USA, 2010.
- 45. Podolyan Y., Leszczynski J. MaSK: A visualization tool for teaching and research in computational chemistry. *Int. J. Ouantum Chem.* 2009. **109**: 8.
- 46. Kulik T., Nastasiienko N., Palianytsia B., Ilchenko M., Larsso M. Catalytic Pyrolysis of Lignin Model Compound (Ferulic Acid) over Alumina: Surface Complexes, Kinetics, and Mechanisms. *Catalysts*. 2021. **11**: 1508.
- 47. 'Swisłocka R. Spectroscopic (FT-IR, FT-Raman, UV absorption,1H and 13 C NMR) and theoretical (in B3LYP/6-311++G** level) studies on alkali metal salts of caffeic acid. *Spectrochim. Acta, Part A.* 2013. **100**: 21.
- Kulyk K, Palianytsia B., Alexander J.D., Azizova L., Borysenko M., Kartel M., Larsson M., Kulik T. Kinetics of valeric acid ketonization and ketenization in catalytic pyrolysis on nanosized SiO₂, γ-Al₂O₃, CeO₂/SiO₂, Al₂O₃/SiO₂ and TiO₂/SiO₂. *Chem. Phys. Chem.* 2017. **18**: 1943.
- 49. Palacios E.G., Juares-Lopes G., Monhemius A.J. Infrared spectroscopy of metal carboxylates: II. Analysis of Fe(III), Ni and Zn carboxylate solutions. *Hydrometallurgy*. 2004. **72**: 139.
- Azizova L.R., Kulik T.V., Palianytsia B.B., Ilchenko M.M., Telbiz G.M., Balu A.M., Tarnavskiy S., Luque R., Roldan A., Kartel M.T. The Role of Surface Complexes in Ketene Formation from Fatty Acids via Pyrolysis over Silica: from Platform Molecules to Waste Biomass. J. Am. Chem. Soc. 2023. 145(49): 26592.
- 51. Bellamy L. Infra-Red Spectra of Complex Molecule. (London: Methuen & Co LTD, 1963).
- 52. Nakanishi K. Infrared Adsorption Spectroscopy (Practical). (San Francisco: Holden Day. Inc. 1962).
- 53. Nastasiienko N., Palianytsia B., Kartel M., Larsson M., Kulik T. Thermal transformation of caffeic acid on the nanoceria surface studied by temperature programmed desorption mass-spectrometry, thermogravimetric analysis and ft–ir spectroscopy. *Colloids Interfaces*. 2019. **3**(1): 34.
- 54. Nastasiienko N., Kulik T., Palianytsia B., Laskin J., Cherniavska T., Kartel M., Larsson M. Catalytic pyrolysis of lignin model compounds (Pyrocatechol, guaiacol, vanillic and ferulic acids) over nanoceria catalyst for biomass conversion. *Appl. Sci.* 2021. **11**(16): 7205.
- González-Baró A.C., Parajón-Costa B.S., Franca C.A., Pis-Diez R. Theoretical and spectroscopic study of vanillic acid. J. Mol. Struct. 2008. 889(1–3): 204.
- 56. Kalinowska M., Piekut J., Bruss A., Follet C., Sienkiewicz-Gromiuk J., Świsłocka R., Rzączyńska Z., Lewandowski W. Spectroscopic (FT-IR, FT-Raman, ¹H, ¹³C NMR, UV/VIS), Thermogravimetric and Antimicrobial

Studies of Ca (II), Mn (II), Cu (II), Zn (II) and Cd (II) Complexes of Ferulic Acid. *Spectrochim. Acta, Part A.* 2014. **122**: 631.

- 57. Sebastian S., Sundaraganesan N., Manoharan S. Molecular structure, spectroscopic studies and first-order molecular hyperpolarizabilities of ferulic acid by density functional study. *Spectrochim. Acta, Part A.* 2009. **74**(2): 312.
- 58. Huang W., Jiang P., Wei C., Zhuang D., Shi J. Low-temperature one-step synthesis of covalently chelated ZnO/dopamine hybrid nanoparticles and their optical properties. *J. Mater. Res.* 2008. **23**: 1946.
- 59. Hachani R., Lowdell M., Birchall M., Hervault A., Mertz D., Begin-Colin S., Thanh N.T.K. Polyol synthesis, functionalisation, and biocompatibility studies of superparamagnetic iron oxide nanoparticles as potential MRI contrast agents. *Nanoscale*. 2016. **8**: 3278.
- 60. Togashi T., Naka T., Asahina S., Sato K., Takami S., Adschiri T. Surfactant-assisted one-pot synthesis of superparamagnetic magnetite nanoparticle clusters with tunable cluster size and magnetic field sensitivity. *Dalton Trans.* 2011. **40**: 1073.
- 61. Nastasiienko N., Kulik T., Palianytsia B., Larsson M., Cherniavska T., Kartel M. Decarboxylation of p-Coumaric Acid during Pyrolysis on the Nanoceria Surface. *Colloids Interfaces*. 2021. **5**(4): 48.
- 62. Kulik T.V., Barvinchenko V.N., Palyanytsya B.B., Lipkovska N.A., Dudik O.O. Thermal transformations of biologically active derivatives of cinnamic acid by TPD MS investigation. *J. Anal. Appl. Pyrolysis*. 2011. **90**(2): 219.
- 63. Ota A., Abramovič H., Abram V., Ulrih N.P. Interactions of p-coumaric, caffeic and ferulic acids and their styrenes with model lipid membranes. *Food Chemistry*. 2011. **125**(4): 1256.
- 64. Dei A., Gatteschi D., Sangregorio C., Sorace L. Quinonoid metal complexes: Toward molecular switches. *Acc. Chem. Res.* 2004. **37**(11): 827.
- 65. Singh V., Naka T., Takami S., Sahraneshin A., Togashi T., Aoki N., Adschiri T. Hydrothermal synthesis of inorganic–organichybrid gadolinium hydroxide nanoclusters with controlled size and morphology. *Dalton Trans.* 2013. **42**: 16176.

Received 16.06.2024, accepted 03.09.2024